**Carbohydrate measurements**

(Adapted from Albalasmeh et al., 2013. Carbohydrate polymers 97: 253-261)

1. Collect cells by centrifugation 4700 rpm for 20 min (culture OD=0.4-0.6);
2. Filter supernatant via 0.2-0.45 μm filter. (NOTE: Filter should be prewashed with ddH2O);
3. Transfer 15 ml of filtered supernatant into 50 ml tube;
4. Add 30 ml acetone and mix by inverting tube 5-6 times;
5. Centrifuge at 4700 rpm for 10 min;
6. Remove upper solution and repeat step 3-5;
7. Remove upper solution and add 5 ml acetone;
8. Centrifuge at 4700 rpm for 5 min and remove upper solution;
9. Dry for 5-10 min;
10. Add 2 ml of hot ddH2O (can use cold ddH2O, but it takes longer to dissolve the pellet); Vortex gently. (Pellet might not dissolve completely, in this case use 2 ml for the step below, transfer everything into a glass tube);
11. Transfer 1 ml of upper solution into a glass tube;
12. Add 3 ml of concentrated sulfuric acid in a test tube and vortex for 10 s ((NOTE: use a glass pipette; vortex very carefully);
13. Cool solution for 5-10 min;
14. Measure absorption at 315 nm in quartz cuvette. Use non-inoculated growth media as a blank (NOTE: non-inoculated growth media control sample should be prepared the same way as supernatant samples).